Original Article

Trends in antimicrobial susceptibility of Gram-negative bacteria isolated from blood in Jakarta from 2002 to 2008

Lucky H. Moehario¹, Enty Tjoa², Ariyani Kiranasari¹, Ika Ningsih¹, Yeva Rosana¹, Anis Karuniawati¹

Abstract

Background: This study examined the susceptibility of Gram-negative bacteria in the bloodstream to antimicrobials with the aim of providing information relevant to the guidance of therapy.

Methodology: Blood specimens received by the Laboratory of Clinical Microbiology, Faculty of Medicine, University of Indonesia, from 2002 to 2008, were analyzed for the presence of Gram-negative bacteria and their susceptibility to four antibiotic groups frequently administered in hospitals and community settings.

Results: During the seven-year period leading up to 2008, approximately 68% of Gram-negative bacteria were identified among all positive isolates from blood specimens. The eight most frequent species found were *Acinetobacter anitratus* (25.8%), *Pseudomonas aeruginosa* (19.5%), *Klebsiella pneumoniae* subsp. pneumoniae (14.5%), *Enterobacter aerogenes* (8%), *Salmonella* Typhi (7.5%), *Escherichia coli* (6.2%), *Alcaligenes faecalis* (5.6%) and *Klebsiella oxytoca* (3.2%). At 80% susceptibility or greater, Ceftriaxone and Cefotaxime were active only on *E. coli* and *S.* Typhi. Cefepime demonstrated activity on all eight species tested except *K. pneumoniae* while Amikacin showed activity against five species, *A. faecalis*, *E. aerogenes*, *E. coli*, *K. pneumoniae* subsp. pneumoniae and *S.* Typhi. Gentamycin was active against three species: *E. aerogenes*, *K. oxytoca* and *S.* Typhi. Ciprofloxacin and Levofloxacin significantly differed in their spectrum: while Ciprofloxacin was active against four of the eight species tested (*E. aerogenes*, *E. coli*, *K. oxytoca*, and *S.* Typhi), Levofloxacin was similar to Cefepime and was active against all eight species except *K. pneumoniae* subsp. pneumoniae.

Conclusions: Since antimicrobials are broadly used in Jakarta, it is important that the information captured in this study be disseminated.

Key words: Gram-negative bacteria, susceptibility to antibiotics

J Infect Dev Ctries 2009; 3(11):843-848.

(Received 25 May 2009 - Accepted 30 October 2009)

Copyright © 2009 Moehario et al. This is an open-access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Introduction

Infections caused by Gram-negative bacteria remain a worldwide problem. Two thirds of septic shock cases are caused by Gram-negative bacteria such as Escherichia coli, Klebsiella, Enterobacter, Proteus, Pseudomonas, and Bacteroides with the remainder being caused by Gram-positive bacteria such as Streptococcus pneumonia, Streptococcus pyogenes, and Staphylococcus aureus [1]. In the United States, Gram-negative organisms were the leading cause of health-care associated blood stream infections before the 1970s, while Gram-positive organisms have since been the predominant microbial isolates [2]. An estimated 200,000 Americans develop Gram-negative sepsis each year with reported mortality rates of 30% to 65% [3]. Gramnegative sepsis is associated with a mortality rate

three times greater than Gram-positive sepsis [4]. In India, Pseudomonas sp. and *E. coli* accounted for approximately 20% of 567 positive isolates derived from patients with fever and sepsis while Citrobacter, Acinetobacter, Proteus, Enterobacter spp. collectively accounted for 80.96% [5].

The present study reported Gram-negative organisms isolated from blood specimens analyzed at the Clinical Microbiology Laboratory of the Faculty of Medicine, University of Indonesia (CML-FMUI), from 2002 to 2008 and their antimicrobial susceptibility patterns. The variation of antimicrobial susceptibility patterns is an important factor in health care awareness associated with managing infectious disease patients and in minimizing the emergence and spread of multidrug-resistant microorganisms.

¹Department of Microbiology, Faculty of Medicine, University of Indonesia, Jalan Pegangsaan Timur 16, Jakarta 10320, Indonesia

²Department of Microbiology, Faculty of Medicine, Catholic University of Atmajaya, Jalan Pluit Raya 2 Jakarta 14440, Indonesia

Materials and methods

Specimens

The present study examined all blood specimens received by the Laboratory of CML-FMUI in Jakarta from hospitals, private practices, or individuals from 1January 2002 to 31 December 2008.

Cultures and antibiotic susceptibility tests

Cultures and susceptibility tests to antibiotics were performed according to CML-FMUI standard practices [6,7] and Performance Standards for Antimicrobial Susceptibility Testing from the Clinical and Laboratory Standards Institute (CLSI) [8]. Cultures were performed using Bactec 9050 (Becton Dickinson) and microorganism identification determined using standard biochemical reactions. The susceptibility of microorganisms to antibiotics was assessed using the disc diffusion method. Antimicrobial susceptibility results were categorized in to three groups: Sensitive (S), Intermediate (I) and Resistant (R) according to CLSI [8]. The antibiotics susceptibility data was then entered into the WHO-Net program.

Antibiotics tested

The following antibiotic discs were included in the present study: Ceftriaxone (30 µg), Cefotaxim (30 µg), Cefepime (30 µg), Levofloxacin (5 µg), Ciprofloxacin (5 µg), Amikacin (30 µg) and Gentamycin (10 µg). Assessments were made based on Gram-negative isolates and numbers of isolates \geq 10. Good activity *in vitro* was defined by antimicrobial susceptibility of 80% or greater for Gram-negative bacteria.

Results

A total of 687 isolates were found in blood specimens analyzed in the targeted 2002 to 2008 time period, with 461 of these isolates being Gramnegative bacteria (67.1%). Eight species of Gramnegative bacteria with a total number of 418 isolates (90.7%) met the present study's criteria. Their antibiotic susceptibility is listed in Table 1. Acinetobacter anitratus, accounting for 25.8% of the isolates, was the predominant species identified during the assessed time period, followed by Pseudomonas aeruginosa at 19.5 %, Klebsiella pneumoniae subsp. pneumoniae at 14.5 Enterobacter aerogenes at 8 %, Salmonella Typhi at 7.5 %, E. coli at 6.2 %, Alcaligenes faecalis at 5.6 %, and Klebsiella oxytoca at 3.2 %.

The spectrum of tested anti-mirobials per type of bacteria is listed in Table 2. Ceftriaxone and Cefotxim was found to be active on only two species of bacteria, *E. coli* and *S.* Typhi. Remarkably, Cefepime and Levofloxacin were found to be active on all eight bacterial species except for *K. pneumoniae*. Amikacin showed good activity on five species of bacteria: *A. faecalis, E. aerogenes, E. coli, K. pneumoniae* and *S.* Typhi. Ciprofloxacin showed activity against four bacterial species: *E.aerogenes, E. coli, K. oxytoca*, and *S.* Typhi. In comparison, Gentamycin was found to be active on only three species of bacteria: *E. aerogenes, K. oxytoca* and *S.* Typhi.

A. anitratus and P. aeruginosa were less susceptible to Ceftriaxone, Cefotaxime, Amikacin, Gentamycin and Ciprofloxacin than to Cefepime and Levofloxacin. A. faecalis showed good sensitivity to three antibiotics, i.e. Cefepime, Amikacin and Levofloxacin, while E. aerogenes exhibited good sensitivity to almost all the antibiotics analyzed, i.e. Cefepime, Amikacin, Gentamycin, Ciprofloxacin, and Levofloxacin. E. coli was sensitive to all antibiotics tested except Gentamycin. K. pneumoniae showed very poor susceptibility to all antibiotics except Amikacin. S. Typhi showed good sensitivity to all antibiotics tested.

Discussion

The management of patients with infectious diseases must focus on patient safety, which includes minimizing the emergence and spread of resistant strains and deciding on the appropriate antimicrobial to administer. Moreover, in sepsis cases, the focal site(s) of infection should be identified to determine which bacteria are involved [9]. In cases with unknown focal site(s) of infection, empiric therapy should be based on the likeliest sources of bacteremia [10].

Ceftriaxone, a third generation Cephalosporin, has broad-spectrum activity against Gram-positive and Gram-negative bacteria. In general, it is considered to be equivalent to Cefotaxime, in terms of safety and efficacy. Ceftriaxone and Cefotaxime are indicated for sepsis syndrome, as well as for pneumonia and skin infection [9]. In our study, Ceftriaxone and Cefotaxim showed less activity with almost all Gram-negative bacteria isolates except for *E. coli* and *S.* Typhi. These results are similar to those from a study conducted in India in which it was found that approximately 50% of fermenter and non-

Table 1. Percentage of eight species Gram-negative bacteria susceptible to antimicrobials in Jakarta from 2002 to 2008.

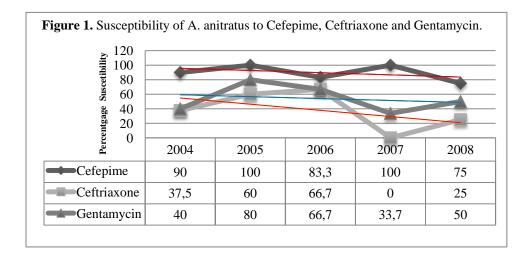
No.	Organisms	No. Isolates	Susceptible to (%)							
			CRO	CTX	FEP	AMK	GEN	CIP	LVX	
1.	Acinetobacter anitratus	119 (25.8%)	50.4	79	89.9	57.1	34.4	68.9	82.3	
2.	Alcaligenes faecalis	26 (5.6%)	27.3	53.8	88.5	87.5	26.9	76	100	
3.	Enterobacter aerogenes	37 (8%)	31.6	68	94.4	100	81.1	91.9	95.8	
4.	Escherichia coli	29 (6.2%)	86.2	90	86.2	82.7	68.9	82.7	82.7	
5.	Klebsiella oxytoca	15 (3.2%)	0	69.2	80.0	NE	86.7	86.7	93.3	
6.	Klebsiella pneumonia subsp. pneumoniae	67 (14.5%)	32.8	46.7	76.1	98.5	50.7	59.7	76.1	
7.	Pseudomonas aeruginosa	90 (19.5%)	42.2	55.1	88.9	56.9	27.8	77.8	92.2	
8.	Salmonella Typhi	35 (7.5%)	92.6	86.7	94.1	100	100	100	100	
	TOTAL	418 (90.7%)								

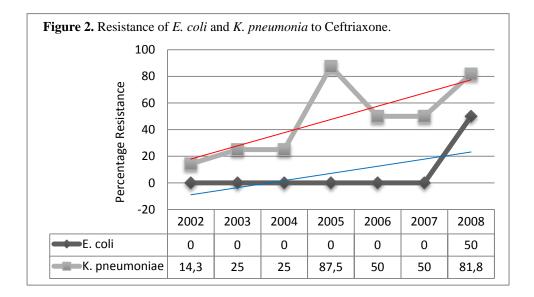
Parentheses refer to percentage of each species to total Gram negative bacteria isolated; CR0: Ceftriaxone, CTX: Cefotaxime, FEP: Cefenime. AMK: Amikacin. GEN: Gentamycin. CIP: Ciprofloxacin. LEV: Levofloxacin: NE: not evaluated

Table 2. Activity of the spectrum of antimicrobials tested on eight species of Gram-negative.

	Organisms		Antibiotics							
No.		CRO	CTX	FEP	AMK	GEN	CIP	LVX		
1.	Acinetobacter anitratus									
2.	Alcaligenes faecalis									
3.	Enterobacter aerogenes									
4.	Escherichia coli									
5.	Klebsiella oxytoca									
6.	Klebsiella pneumoniae subsp. pneumoniae									
7.	Pseudomonas aeruginosa									
8.	Salmonella Typhi									

Black dots indicate good activity in vitro was defined as 80% or greater susceptibility of Gram-negative





fermenter Enterobacteriaceae were susceptible to those antimicrobials [5]. In Europe, Canada and the United States in the years 2000 and 2001, *E. coli* and *K. pneumonia* exhibited approximately 90% susceptibility to both Ceftriaxone and Cefotaxim, while *Enterobacter spp.* demonstrated a lower susceptibility [9].

The present study showed that *P. aeruginosa* is intrinsically resistant to Ceftiaxone, Cefotaxim, Kanamycin and Neomycin. Similar to our results, *P. aeruginosa* showed less than 55% susceptibility to those antimicrobials in another study [9]. In European countries, Amikacin was active *in vitro* against *P. aeruginosa* (> 80% susceptibility) [9]; however, this was not so in India [5] and Indonesia (as found in the present study). Other commonly used Aminoglycoside antibiotics such as Gentamycin showed good activity against Gram-negative bacteria

in Europe, but exhibited only 48% to Enterobacteriaceae in India. The present study found that more than 80% of *E. aerogenes*, *K. oxyyoca* and *S.* Typhi were susceptible to Gentamycin. Notably, Aminoglycoside has been found to be ineffective *in vivo* against *S.* Typhi despite its activity *in vitro*; therefore, it is not recommended for treatment of *S.* Typhi infections such as Typhoid Fever [8].

Cefepime is a fourth-generation Cephalosporin antibiotic which has an extended spectrum of activity against both Gram-positive and Gram-negative bacteria and has greater activity than third-generation agents. Cefepime is usually reserved for infections caused by *P. aeruginosa* and multidrug-resistant microorganisms. It is an effective antimicrobial of choice for critically-ill sepsis patients as well as patients with febrile neutropenia. Several studies have shown that Cefepime is associated with low

mortality rates, rapid clinical improvement, minimal drug resistance, and minimal adverse effects in patients with severe sepsis [12]. At present, Cefepime demonstrates good activity against Gramnegative bacteria *in vitro* including *P. aeruginosa*, which is consistent with the findings of the study. However there are issues surrounding the clinical safety of Cefepime as it has been associated with an increased mortality rate when used for different types of infections [13]. As a result, the administration of Cefepime is becoming less common.

Monotherapy has been associated with clinical failure in cases with serious infections caused by P. aeruginosa. For that reason, a combination treatment with beta lactam antimicrobials and Fluoroquinolone or Aminoglycoside should be considered [8]. In comparison to Ciprofloxacin, Levofloxacin exhibited superior activity in Gram-negative bloodstream isolates (see Table 2). Levofloxacin was active against almost all the Gram-negative bacteria assessed in this study, including P. aeruginosa. Ciprofloxacin did not exhibit similar activity. The results of the present study are consistent with those of Flynn et al. [14] which showed that the minimum inhibitory concentrations (MICs) for Levofloxacin against multi-resistant P. aeruginosa strains were generally two- to fourfold higher than those of Ciprofloxacin. S. Typhi exhibited susceptibility to all the antibiotics tested; however, due to the low numbers of isolates tested in the present study, it is possible that antimicrobialresistant S. Typhi strains were not found. It is not recommended that Aminoglycoside be used in treating Salmonella and Shigella infections as in vitro results did not correlate with in vivo conditions, and thus it was not clinically effective [8]. During the seven-year course of the present study, A. anitratus the most commonly found isolates. Acinetobacter species are commonly recognized as commensal bacteria, but they have occasionally caused hospital infections. This microorganism was frequently found to be resistant to antimicrobial agents, and thus infections caused by Acinetobacter species were difficult to treat. In the present study A. anitratus demonstrated a decreasing susceptibility trend to third- and fourth-generation Cephalosporine antimicrobials (see Figure 1). Klebsiella species and E. coli were predominantly extended-spectrum beta lactamase (ESBL) producers, making microorganisms clinically resistant to penicillins, Cephalosporines or Aztreonam despite apparent in vitro susceptibility. Results from the present study indicated an increasing number of K. pneumonia and E. coli strains resistant to Ceftriaxone from 2002 to In conclusion, Cefepime 2008 (see Figure 2). demonstrated superior in vitro activity for Gramnegative bacteria of the bloodstream compared to Ceftriaxone and Cefotaxim. Compared Ciprofloxacin, Levofloxacin showed good activity against almost all the types of microorganisms found. Generally, Levofloxacin and Cefepime demonstrated good activity against both fermenter and nonfermenter Enterobacteriaceae. Since antibiotic susceptibility information is important in determining the guidelines for antibiotic therapy of infectious diseases, it is important that this information be collected on a regular basis.

References

- Lippincott, Williams & Wilkins (2005) Professional guide to disease, 8th edition. Ed: Etkin S, Lenker D, Mils E. Ambler, PA, Springhouse, pp. 187-188.
- Albrecht SJ, Fishman NO, Kitchen J, Nachamkin I, Bilker WB, Hoegg C, Samel C, Barbagallo S, Arentzen J, Lautenbach E (2006) Reemergence of Gram-negative healthcare—associated bloodstream infections. Arch Intern Med 166: 1289-1294.
- Angus DC (2000) E5 murine monoclonal antiendotoxin antibody in Gram-negative sepsis. JAMA 283: 1723-1730.
- Seneca H and Grant JP (1976) Gram-negative sepsis and endotoxic shock. Antibiotics and Chemotherapy 21: 52-61.
- Mehta M, Dutta P, Gupta V (2005) Antimicrobial susceptibility pattern of blood isolates from a teaching hospital in North India. J Infect Dis 58: 174-176.
- Standard Operating Procedure for Clinical Microbiology Examination (1999) Laboratory of Microbiology of Department Microbiology, Fac. of Medicine Univ. Indonesia, Jakarta.
- 7. Standard Operating Procedure for Clinical Microbiology Examination (2004) Laboratory of Microbiology of Department Microbiology, Fac. of Medicine, Univ. Indonesia, Jakarta.
- 8. Performance Standards for Clinical Laboratory Standard Institute (CLSI), year up to date.
- Wenzel RP, Sahm DF, Thornsberry C, Draghi DC, Jones ME, Karlowsky JA (2003) In vitro susceptibilities of Gramnegative bacteria isolated from hospitalized patients in four European countries, Canada, and the United States in 2000-2001 to expanded-spectrum Cephalosporins and comparator antimicrobials: Implications for therapy. Antimicrobial Agents and Chemotherapy 47: 3089-3098.
- 10. Evans M (2007) Antibiotic guidelines for the empirical treatment of sepsis in immune competent adult. Nottingham Antibiotic Guidelines Committee.
- 11. Leclercq R, Canton R, Giske C, Heisig P, Rossolini GM, Winstanley T (2008) Expert rules in antimicrobial susceptibility testing. European Committee on Antimicrobial Susceptibility Testing (EUCAST).
- 12. Stevanov C, Doikov I, Dimov R, Deenichin G, Delikostadinov G (2002) Clinical and bacteriological result from the administration of Cefepime (Maxipime) in patients with severe sepsis. Med Sci Monitoring 8: 197-102.

- 13. Sonck J, Laureys G and Verbeelen D. (2008) The neurotoxicity and safety of treatment with Cefepime in patients with renal failure. Nephrology Dialysis Transplantation 23: 966-970.
- Flynn CM, Johnson DM, Jones RN (1996) In vitro efficacy of Levofloxacin alone or in combination tested against multi-resistant *Pseudomonas aeruginosa* strains. J of Chemo 8: 411-415.

Conflict of Interest: No conflict of interest is declared.

Corresponding Author

Lucky H. Moehario

Department of Microbiology, Faculty of Medicine, University of Indonesia

Jalan Pegangsaan Timur no. 16, Jakarta 10320, Indonesia Email: luckyhmoehario@gmail.com