Preliminary remarks on assembly whole genome sequencing of MDR M. tuberculosis isolated in Vietnam

Ngo Viet Quynh Tram¹, Nguyen Hoang Bach¹, Nguyen Thi Chau Anh¹, Huynh Hai Duong¹, Le Nu Xuan Thanh¹, Le Van An¹, Stefano Ferroni¹, Piero Cappuccinelli^{1,2}

¹Carlo Urbani Centre, Dept. of Microbiology, Hue College of Medicine and Pharmacy, Vietnam

Key words: MDR M. tuberculosis; genome sequencing; Vietnam

J Infect Dev Ctries 2012; 6(1):95-96.

(Received and Accepted 14 December 2011)

Copyright © 2012 Tram *et al.* This is an open-access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Tuberculosis remains a major cause of morbidity and mortality in many countries and a significant public health problem worldwide. The emergence of drug resistant strains and particularly multidrug-resistant strains of *Mycobacterium tuberculosis* has become a significant public health problem in a number of countries and an obstacle for an effective control of tuberculosis.

Vietnam is a high-burden country for tuberculosis [1]. In Vietnam, almost 30,000 people die every year from TB (one death every 18 minutes). Vietnam, with its estimated 175,000 new cases per year, ranks 12th among the 22 countries that bear 80% of the global TB burden [1,2]. If TB is detected early and treated properly by using a combination treatment for six to nine months, the patients quickly become non-infectious and are eventually cured. Multi-drug resistant TB (MDR-TB) and extensively drug-resistant TB (XDR-TB), HIVassociated TB, and weak health systems are major challenges in Vietnam. There are an estimated 7,000 new MDR-TB cases and 6,400 new TB/HIV cases every year. Both of these forms of complicated TB carry a high risk of early mortality [1]. Although tuberculosis is still a public health problem in Vietnam, there is little information about the genetic characteristics of the isolates. A better knowledge of the molecular characteristics of M. tuberculosis strains will contribute to our understanding of the transmission dynamics of the disease within the country.

One strain of *M. tuberculosis* (MTB_HUE_20 strain), which was isolated from a smear-positive sputum specimen of a 47-year-old male patient with

typical clinical features of new tuberculosis in Hue, Vietnam, was resistant to isoniazid and rifampicin (multidrug resistant), positive with IS6110 PCR. Furthermore, its spoligotyping pattern did not match any described genotype in the SpolDB4 database (unknown genotype). This strain was chosen for resequencing of the whole genome based on paired-end sequencing on the Illumina GAIIx platform (Illumina Inc, San Diego, CA, USA). Thirty micrograms of the genomic DNA was purified by the CTAB method and sent to BaseClear Co DNA Sequencing Service in the Netherlands to perform the sequencing. De Novo genomic assembly was performed with the Velvet program (EMBL-EBI, Hinxton, Cambridge, UK) [3]. Gene prediction and translate predicted genes into proteins with GeneMarkS [4]. All ORFs obtained were annotated with the Blast2Go program [blast2go] (BioBam Bioinformatics S.L., Valencia, Spain), and GFF files were created to annotate the circular genome MTB HUE 20 strain with Geneious software (Biomatters Ltd, Auckland, New Zealand) [5].

Some preliminary observations on the assembled whole genome sequencing of MTB_HUE_20 strain were as follows: the full length is 4,397,928 bp, less than that of H37Rv strain of about 14Kb; percentages of A, T, C, G and high percentages of G and C are similar to those of H37Rv strain; this strain harbors only one copy of IS6110 and has 2329 SNP (single-nucleotide polymorphisms), of which 1257 lead to change of amino acid and 159 DIP (Deletion Insertion Polymorphisms), of which 105 lead to change of amino acid when compared to whole genome of

²Department of Biomedical Sciences, University of Sassari, Italy

H37Rv strain; the lack of 15 copies of IS6110 159 DIPs cause the length combining MTB_HUE_20 strain shorter than H37Rv strain; interestingly this multidrug resistant strain has a mutation in the katG gene but no mutation in the rpoB gene.. Mutations in DNA-dependent RNA polymerase (rpoB) gene is among the most frequent in RIFresistant strain [6] and catalase-peroxidase (katG) gene in INH-resistant strain [6,7]. Previous studies indicated that less than 5% of resistant strains do not show a mutation in the *rpoB* resistance region [7,8] and rare inconsistencies of mutated M. tuberculosis strains, one of them may be clinically resistant, which could suggest other mechanism for resistance (9]. Maybe the MTB HUE 20 strain is one of these strains.

This report confirms a multidrug resistant *M. tuberculosis* strain with KatG mutation. Multidrug resistant tuberculosis could become an emerging problem in Vietnam; therefore early detection of drug resistance and proper treatment are needed for the effective control of drug resistant tuberculosis.

References

- WHO (2005) Global tuberculosis control: surveillance, planning, financing. WHO report Geneva: World Health Organization.
 - http://allafrica.com/download/resource/main/main/idatcs/0001 0603:4e04b1cdf15c013b0bf2117aafb1eee6.pdf. Last accessed 31 December 2011.
- URC (2011) Vietnam Improving TB Detection and TB-HIV Services Integration. Available: http://www.wpro.who.int/vietnam/sites/dcc/tb/. Last accessed 31 December 2011.

- Zerbino DR and Birney E (2008) Velvet algorithms for de novo short read assembly using de Bruijn graphs. Genome Research 18: 821-829.
- Besemer J, Lomsadze A, Borodovsky M (2001) GeneMarkS: a self-training method for prediction of gene starts in microbial genomes. Implications for finding sequence motifs in regulatory regions. Nucleic Acids Research 29: 2607-2018.
- Drummond AJ, Ashton B, Buxton S, Cheung M, Cooper A, Heled J, Kearse M, Moir R, Stones-Havas S, Sturrock S, Thierer T, Wilson A (2010) Geneiousv5.1. Available: http://www.geneious.com. Last accessed 10 December 2011
- Zhang M, Yue J, Yang YP, Zhang HM, Lei JQ, Jin RL, Zhang XL, HH Wang. (2005). Detection of mutations associated with isoniazid resistance in *Mycobacterium* tuberculosis isolates from China. J Clin Microbiol 43: 5477-548
- Riska PF, Jacobs WR, Alland D (2000) Molecular determinants of drug resistance in tuberculosis. Int J Tuberc Lung Dis 4: S4-10.
- Mani C, Selvakumar N, Narayanan S, Narayanan PR (2001) Mutations in the rpoB gene of multidrug-resistant Mycobacterium tuberculosis clinical isolates from India. J Clin Microbiol 39: 2987-2990.
- Pierre-AudigierC and GicquelB (2010) The contribution of molecular biology in diagnosing tuberculosis and detecting antibiotic resistance. Available: http://www.eumednettb.org/doc/Revue.pdf. Last accessed 10 December 2011.

Corresponding author

Ngo Viet Quynh Tram Carlo Urbani Centre Department of Microbiology Hue College of Medicine and Pharmacy 06 Ngo Quyen, Hue, Vietnam Telephone / Fax: 0084 54 3837447 Email: qtramny@gmail.com

Conflict of interests: No conflict of interests is declared.